Impact of improved hygiene: Farrowing accommodation and liquid feeding systems

Peadar Lawlor¹, Keely Halpin^{1,2}, James Cullen^{1,2}, Florence Viard^{1,3}, John O'Doherty³ and Gillian Gardiner²

¹Teagasc, Moorepark, ²South East Technological University, Waterford and ³University College Dublin, Dublin







Introduction

- ↑ internal biosecurity: ↑ pig growth and ↓ mortality & antibiotic usage
- But, many factors are associated with internal biosecurity
- Impact of measures such as cleaning & disinfection routines not always clear
 - Implementing correctly takes time, and
 - Temptation to take short cuts or, worse, avoid altogether, particularly where labour & space limiting
- Two very different but critically important areas on the unit regarding hygiene
 - Farrowing accommodation
 - Liquid feeding systems



1. Farrowing Accommodation Hygiene







Internal Biosecurity- Pig Health Check





Introduction

- High use of antibiotics linked to spread of AMR from animals to humans
 - Increased restrictions on antibiotic use in EU January 2022
 - Therapeutic levels of in-feed zinc oxide banned in EU June 2022

All happening when litter size \uparrow - piglet weight and health to weaning

- Internal biosecurity measures shown to ↑ pig growth, ↓ mortality (Laanen, et al. 2013) and ↓ antibiotic usage (Postma, et al. 2017)
- We believe farrowing accommodation hygiene to be particularly important
- Objective: ↓ the need to medicate suckling piglets & ↑ piglet growth by implementing an optimised sanitisation routine in farrowing accommodation





Farrowing Accommodation Hygiene

Sub-optimal vs. improved/optimal cleaning and disinfection protocol

- ~22 litters/pens on each protocol
- Average born alive 14.9
- 2 batches of pigs

Parameters measured:

- Microbiology: Total bacterial and *Enterobacteriaceae* counts in farrowing pens
- Growth: Individual piglet weight
- Health: Clinical cases, no. injections, antibiotic & anti-inflammatory usage per litter



Study

Optimal sanitisation routine for farrowing accommodation



Pre–soak pens with water overnight (≤18hr)



Chlorocresol-based disinfectant (Interkokask®) Dry 6 days, blow heater 1st 24 hr



Detergent (Blast Off - Carboxylic acid) - 20 min Power wash Dry overnight with blow heater



Sows: washed & disinfected (Virkon® S – potassium sulfate) pre-entry to farrowing crates

Sub-optimal sanitisation routine for farrowing accommodation



Washing pens with water

Dry overnight (≤18 hr)



Swabbing of farrowing pens

Areas swabbed



Sow feeder Swabbing of pens • 1) Before washing (pens containing organic matter) **Piglet lying area** • 2) After disinfectant application (2 hrs) • 3) 24 h after disinfectant application Floor area behind

- 4) 72 h after disinfectant application
- 5) After drying / as sows enter • farrowing crates

Wall behind the SOW

the sow

Piglet drinker

Sow's udder



Results - Total Bacterial Counts



Optimal



¹ Detection limit for floor area behind sow before washing (1.4 Log CFU/cm²)
² Detection limit for floor area behind sow after washing (0.4 Log CFU/cm²)



¹ Detection limit for wall behind the sow before washing (1.4 Log CFU/cm²) ² Detection limit for wall behind the sow after washing (0.4 Log CFU/cm²)



Results

Pre-weaning pig growth performance and therapeutic treatments

17

15

13

9

5

3

% 11



6 P < 0.001 5 Injections / Litter - 79% 4 3 0

No. injections / litter

Antibiotic usage / litter

Mortality

P = 0.22

- 28%





No. of clinical cases / litter



Anti -inflammatory usage / litter



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Optimal

Implications

- - Optimal; 32 min per pen, 4 steps, 6 days drying (3 days drying is sufficient)
 - Sub-optimal; 23.5 min per pen, 2 steps, overnight drying.
- Requires sufficient farrowing accommodation to implement properly
- Sub-optimal may seem basic
 - Compared to sanitisation regime in Moorepark at the time, yielded similar results
 - Representative of commercial sanitisation regimes
- A little more time and effort yields dramatic benefits in terms of reducing medication usage and increasing piglet growth





2. Liquid Feeding System Hygiene









Introduction

- No standard protocol to optimise liquid feeding system hygiene
- Poor hygiene: proliferation of undesirable bacteria and fungi
 - Loss of energy and amino acids from the feed
 - Poorer Feed Efficiency and potentially reduced growth
- Objective: Test a practical & easy to apply feeding system sanitisation protocol
 - Remove/disrupt biofilms in the pipes and mixing tank
 - ✓ Suppress *Enterobacteriaceae* and yeast & mould growth in liquid feed
- Combination of an alkali wash followed by an acid rinse



Liquid Feeding System Hygiene Protocol



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Liquid Feeding System Hygiene Protocol

Before cleaning

After cleaning

Mixing tank lid



Day -1

Physical cleaning (wash & scrub)



Wash balls and exhaust pipe



Mixing tank scrubbed and power washed



Results: Mix tank swab



Results: Pipe swab



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Results: Scanning electron microscopy



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Results

Group E. coli Enterobacteriaceae Lactic acid bacteria Yeasts Total aerobic count Moulds



Fresh trough feed



- Feed-associated microbes proliferate in feed despite improved system hygiene
- Should we be surprised?
- ADFI: 2,854 g/d; ADG: 1,216 g/d; FCE: 2.35





Conclusions & Implications

- Improved hygiene of mixing tank and pipes
 - Opportunity to control/reduce re-colonisation of system
- Pipe biofilm not completely removed but $\downarrow E. coli, Enterobacteriaceae & moulds$
 - Implications for pathogen presence & mycotoxin production
- Little impact of cleaning on feed microbial composition
 - Focus on feed! Acidify feed / use of homofermentative inoculants
 - Control/reduce microbial load of feed + good water quality
 - Good system hygiene will prevent recolonization of feed mix



Acknowledgements - PigNutriStrat

Funding

The PigNutriStrat project is funded by the Irish Department of Agriculture, Food and the Marine's Competitive Research Funding Programmes (Grant no: 2019R518).







Acknowledgments – WetFeed-2

Partners













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Funders









Results

Dry feed



(n = 3)

Water

(n = 2)

Dry feed on a liquid feed basis



